
BREAKING UP DORMANCY OF ADANSONIA DIGITATA L. SEEDS AND REGENERATION OF PLANTLETS FROM STEM NODAL SEGMENTS IN VITRO

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ABSTRACT

*In Yemen, there are only three trees of *Adansonia digitata* L. which is adopted in this study. Natural regeneration and independent germination of *Adansonia digitata* is poor although it is one of the tallest trees in the world that was recorded to live about 4000 years. Unfortunately, the tree is a subject of extinction. In the present study, the seeds of *Adansonia digitata* L. were pretreated with full and half strength of hydrochloric, sulfuric and nitric acids for one and/or two hour as a mean of scarification to break the dormancy of the seeds. Seeds were then surface sterilized and allowed to germinate on hormone-free MS basal salts culture medium in vitro. The obtained seedlings were used as a source of nodal segments explants for propagation in vitro. Results of the present study have shown that all of the acid treatments overcame seed dormancy and enhanced seed germination in various degrees. The percentage of seed germination increased from 18.6% in the control group to 86.6% using a pretreatment of half strength sulfuric acid for one hour. The germinated seeds grown into seedlings of a mean of 11.1 centimeter after 8 weeks of growth. Stem nodal segments were cultured on agar solidified MS basal salts supplemented with various concentrations of benzyle adenine (BA) and naphthalene acetic acid (NAA). The maximum regenerant shoot length (15.1±0.3 cm) was obtained with 8 mg/l BA plus 0.5 mg/l NAA while The maximum number of leaves per regenerant reached up (7.2±0.2) using 10 mg/l BA + 1.0 mg/l NAA. The obtained regenerants developed roots on the same culture media. They were acclimatized for one week and then transferred to soil.*

*Keywords: *Adansonia digitata* L, Seed dormancy, Regeneration, In vitro.*

INTRODUCTION

Adansonia digitata L. (Bombacaceae), native to Africa, is one of the tallest trees in the world and may live for 4000 years. *Adansonia* is regarded as the “Queen of all carbon storage trees” as the tender roots, tubers, twigs, fruits, seeds, leaves and flowers are all edible and they are common ingredients in traditional dishes in rural areas in Africa (Sundarambal et al., 2015). In its African natural habitats, *Adansonia digitata* is considered an economically important tree species for rural people and also on the national level because of its medicinal (Kamatou and Vermaak Vilijoan, 2011) and nutritional (Zahrau et al., 2014) uses as well as benefits of the tree as a source of raw material for many other purposes (for example the bark of the tree is used as an anti fire fiber). From various parts of the plant, reasonable amounts of phytochemically active metabolites like steroids, flavonoids, epicatechin, campesterol, Tocopherol, adansonin, amino acids, vitamins and minerals. (Suganth et al., 2014; Jitin et al.,

2015) has been detected. This may explain why It has been used effectively in the treatment of bronchial asthma, dermatitis, sickle cell anemia, as a diuretic, anti-diabetic, anti-oxidant, anti inflammatory agent, antidote for poison, antibacterial agent, anti viral, anti-trypanosome, as a laxative and also used against diarrhea and dysentery (Donaties et al., 2011; Singh et al., 2013).

Adansonia digitata (which is locally called “Shagarat Al-Ghareeb” in Yemen) is not an endemic tree. The total number of trees found in Yemen by botanists is just 3 (personal communication). There are suggestions that it was occasionally brought up from its natural African habitats to Yemen by Arab traders (Burton, 1969). Under the circumstances of Yemen, *Adansonia digitata* exhibits many problems with regards to flowering, fruiting and accordingly propagation. In India, The tree at present is also facing a crisis of survival and is enlisted as an endangered species in the Red data book with only 30 to 40 trees available (Singh et al.,

2010). In Senegal, N'Doye et al., (2012) also reported that the tree is now threatened with depletion and worse extinction. Gebauer, et al., (2002) reported that *Adansonia digitata* suffers from high risk of extinction because of the lack of its natural regeneration and hence, practical ex situ conservation measures are urgently needed to preserve genetic diversity and maintain multiple specimens. Danthu et al., (1995) reported that Baobab (*A. digitata*) seeds have very hard seed coats and germination is usually less than 20%. Dormancy of seeds of *Adansonia digitata* can be attributed partly to the testa and partly to the pulp. According to Falemara et al., (2013), the restricting factor in germination is due to the fact that the seed coat is impermeable to water. Cultivation of baobab necessitates that the seeds be pre-treated to enhance the accessibility of water and oxygen into the seeds before planting in order to break dormancy and to obtain optimum germination and improve performance for plantation establishment. Several methods, such as wet heat treatment, total or partial seed decoating and scarification of seeds with concentrated acids, herbicides, fungicides and growth regulators has been used to break up seed prmancy.

In vitro regeneration of *Adansonia digitata* plants via seed culture was carried out by Katsuki and Sie (2007), Singh et al., (2010) and also from different explants of seedlings by N'Doye et al., (2012). The main objective of the present study is to break up the dormancy of seeds by acid pretreatment and then use the nodal segments of the obtained seedlings as explants for in vitro regeneration.

MATERIALS AND METHODS

Plant material: The seeds of *A. digitata* L. were isolated from fruits brought from Sudan. The seeds were subjected to viability test through the floatation method. This is based on the observation that sound seeds of most spe-

cies sink in water while empty, defective and dead seeds float.

Scarification of seeds: This was carried out by immersing the seeds in full and half strength of hydrochloric acid, sulfuric acid and nitric acid for one or two hours. The seeds were then washed with distilled water several times to get rid of any traces of the acid treatments before in vitro germination via seed culture.

Surface sterilization of seeds: This was carried out according to Chawla, (2003) by submerging the seeds into a solution of 70% ethanol with continuous and gentle steering for one minute, transferring them to 100ml conical flask containing 20% solution of commercial sodium hypochlorite (1% active chlorine) with continuous gentle steering for 15 minutes. The sterilant decanted and the seeds washed with 3 successive rinses of sterile distilled water under aseptic conditions. The seeds were dried between two layers of sterile filter papers in a Petri-dish.

Germination of seeds: This was carried out under aseptic conditions by sawing the seeds on the surface of hormone – free Murashige and Skoog's (1962) basal salts media solidified with 0.8% agar. Each culture tube contained 30 ml of culture media and one seed. Germination was allowed to take place at room temperature.

Regeneration : The obtained seedlings from germination experiment were used as sources of stem nodal segments. The nodal segments were aseptically cultured on MS culture media supplemented with BA (0, 2, 4, 6, 8, 10 mg/l in combination with NAA 0.0, 0.5, 1.0 mg/l). The cultures were kept at room temperature, photoperiod of 16 hours light, 8 hours dark.

Statistical analysis

Germination of seeds and response of nodal segments to various hormonal treatments was calculated as %. Shoot length and number of leaves per regenernt was calculated as a mean of 10 readings \pm standard error.

Table (1): Effect of acid pretreatments and soaking periods on germination of *Adansonia digitata* seed germination

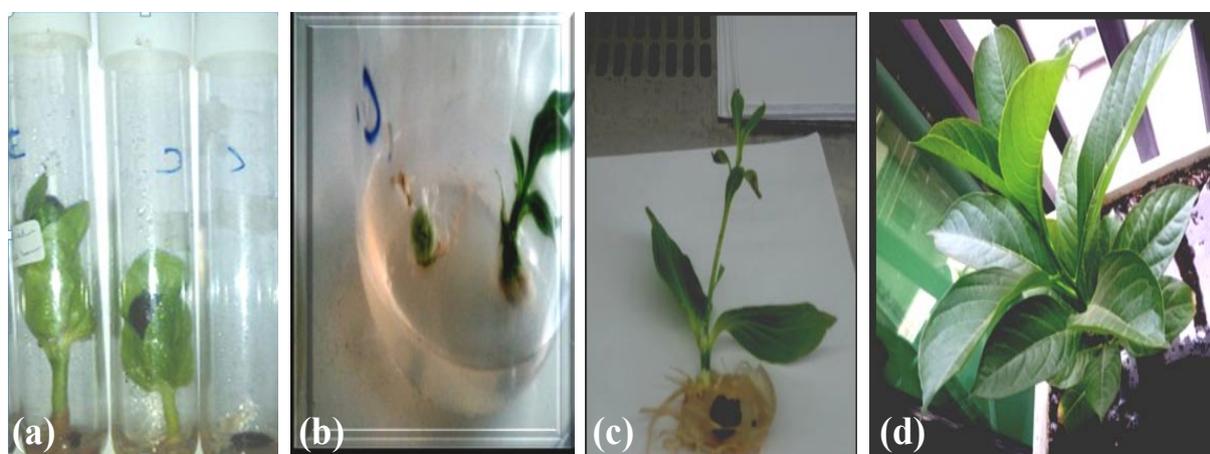
| Treatment | | Control | Hydrochloric | | | | Sulfuric | | | | Nitric | | | | |
|---|---------|---------|---------------|------|--------------|------|---------------|------|--------------|------|---------------|------|--------------|-----|--|
| | | | Full strength | | 1/2 Strength | | Full strength | | 1/2 strength | | Full strength | | 1/2 strength | | |
| Strength | | | I | II | I | II | I | II | I | II | I | II | I | II | |
| Period (hrs) | | | | | | | | | | | | | | | |
| No. of germinated seeds After a period of | 10 days | 3 | 3 | 7 | 12 | 5 | 9 | 20 | 18 | 12 | 4 | 6 | 4 | 6 | |
| | 20 days | 6 | 18 | 21 | 23 | 22 | 14 | 12 | 34 | 11 | 25 | 13 | 16 | 12 | |
| | 30 days | 2 | 4 | 0 | 1 | 0 | 8 | 4 | 0 | 2 | 1 | 0 | 0 | 0 | |
| Total No. of germinated seeds per 60 | | 11 | 25 | 28 | 36 | 27 | 31 | 36 | 52 | 25 | 30 | 29 | 20 | 18 | |
| % of germination | | 18.3 | 41.6 | 46.6 | 60.0 | 45.0 | 51.6 | 60.0 | 86.6 | 41.6 | 50.0 | 48.3 | 33.3 | 30. | |

Table (2): Shoot length of *A. digitata* seedlings germinated in vitro on MS hormone free culture medium

| | | | | |
|------------------------|-----|-----|-----|------|
| Growth period in weeks | 2 | 4 | 8 | 12 |
| Shoot length in cm. | 4.3 | 6.1 | 8.5 | 11.1 |

Table (3): Effect of various hormonal concentrations on number of nodal segments which showed bud development, height of regenerants developed (cm) and number of leaves per regenerant. Each value is a mean of 10 readings \pm standard error. Measurements were taken after one month of growth on MS.

| NAA mg/l. | | BA mg/l. | | | | | |
|-----------|-------------------------------|----------|---------------|----------------|----------------|----------------|----------------|
| | | 0 | 2 | 4 | 6 | 8 | 10 |
| 0 | No., of explants responded/10 | 1/10 | 0/10 | 4/10 | 4/10 | 8/10 | 8/10 |
| | Regenerant height (cm) | | | 6.1 \pm 0.1 | 8.2 \pm 0.3 | 10.1 \pm 0.2 | 13.8 \pm 0.3 |
| | Number of leaves | ... | | 3.0 \pm 0.3 | 3.3 \pm 0.2 | 5.2 \pm 0.3 | 6.1 \pm 0.5 |
| 0.5 | No., of explants responded/10 | 0/10 | 2/10 | 4/10 | 4/10 | 8/10 | 8/10 |
| | Regenerant height (cm) | | 4.5 \pm 0.2 | 11.1 \pm 0.6 | 10.8 \pm 0.5 | 15.1 \pm 0.3 | 14.3 \pm 0.5 |
| | Number of leaves | ... | 2.9 \pm 0.1 | 5.3 \pm 0.3 | 5.9 \pm 0.1 | 8.8 \pm 0.5 | 7.2 \pm 0.2 |
| 1.0 | No., of explants responded/10 | 0/10 | 0/10 | 4/10 | 6/10 | 6/10 | 10/10 |
| | Regenerant height (cm) | | 0 | 6.8 \pm 0.1 | 13.3 \pm 0.2 | 13.5 \pm 0.3 | 14.3 \pm 0.2 |
| | Number of leaves | | 0 | 3.7 \pm 0.4 | 8.2 \pm 0.3 | 6.3 \pm 0.5 | 7.2 \pm 0.2 |

Figure (1): Regeneration of *A. digitata* in vitro. (a) Germination of seeds on MS, (b) Lateral bud growth from nodal segments, (c) Rooted plantlets, (d) Acclimatized potted plant.

RESULT AND DISCUSSION

From the results of the present study (table 1) that the percentage of germination of control group seeds was 18.3%. This result may agree and confirm the work done by Danthu et al., (1995) who reported that Baobab seeds have very hard seed coats and germination is usually less than 20%. Esenowo, (1991), also asserted that dormancy in seeds of *Adansonia digitata* (L.) can be attributed partly to the testa and partly to the pulp. Falemara et al., (2013) reported that the restricting factor in germination is due to the fact that the seed coat is impermeable to water and thus, the cultivation of baobab seeds necessitates a pre-treatment step to enhance the accessibility of water and oxygen into the seeds before planting, in order to break dormancy .

Results obtained in this study (table 1) shows that all acid pretreatments treatments could break up seed dormancy and enhance seed germination over the control group seeds. The best percentage of seed germination reached up 86 % in response to treatment with half strength sulphuric acid for one hour, a result which may agree with the results obtained by Esenowo, (1991) who reported that, the most effective method of *A. digitata* seed pretreatment was scarification with H₂SO₄ where the germination percentage reached up to 98%. In another study carried out by Danthu, et al., (1995), it was found that the treatment of baobab seeds with concentrated sulphuric acid for 6-12 hrs led to germination of more than 90% of the seeds within twenty days of culturing , a result which may be also more or less similar to the results obtained in this study. It is to be remembered also that sulphuric acid pretreatment of *Azizelia* African seeds reduced the dormancy period and yielded a more uniform and regular germination by Amusa (2011). Results of the present study (table2) shows that the germinated seeds could grow to reach up a height of 11.1 cm after 12 weeks on hormone free MS culture media which was renewed every 4 weeks.

The results obtained in this study, as illustrated in table (3), have shown that the development of lateral buds into shoots needed the presence of benzyl adenine as a must was directly affected by

the cytokinin/auxin ratio in the culture medium. The concentration of 10 mg/l of benzyl adenine in combination with 1 mg/l of NAA resulted in a response of 100 percent bud development where all the tested nodal segments explants produced shoots from their lateral buds. The number of leaves per single shoot ranged from 2.9±0.1 (using 2 mg/l BA + 0.5 mg/l NAA) to 7.2±0.2 (using 10 mg/l BA + 1.0 mg/l NAA). The maximum regenerant height (15.1±03 cm) was obtained with 8 mg/l BA plus 0.5 mg/l NAA. The obtained regenerants directly developed root morphogenesis without any need to transfer them to a rooting medium. This probably due to the presence of auxin in the shoot growth media used. The first attempt of regeneration of *A. digitata* from stem nodal segments were carried out by Katsuaki and Sie (2007) where they reported that Shoots were obtained from nodal segments of in vitro germinated *A. digitata* cultured on 1/2LP medium containing 10 µM BAP and that rooted plantlet was regenerated on half-strength woody plant medium containing 3.5 µM IBA and 0.32 µM NAA for the first time. Successful regeneration trial was also done by N'Doye et al., (2012) from different explants of seedlings and Rolli et al., (2014) via enhancing axillary bud multiplication. Rolli et al., (2014) In vitro shoot multiplication was achieved by enhanced axillary bud proliferation of sterilized two-node segments. Bud break was dependent on cytokinin supply (a result that was also clearly observed in the present study) but the combination of 1.0 or 10.0 µM zeatin riboside and 10.0 µM indole-3-butyric acid (IBA) increased the formation of microshoots after 8 weeks of culture. Regenerated microshoots rooted successfully in in vitro nutrient medium containing 10.0 µM IBA and normally grew in a greenhouse after acclimatization. Some of the steps of the regeneration process of *A. digitata* can be seen in figure (1).

CONCLUSION

Adansonia digitata L. is an endangered plant not only in the countries where it was introduced in but also in its natural habitat due to overharvesting, high level of seed dormancy and poor regeneration and slow growth. In Yemen, there are only three trees and in all India just 30-40

trees. *Adansonia digitata* (which is called by the public in Yemen “Shagart El-Ghareeb”) is one of the tallest trees in the world that was recorded to live about 4000 years. There are many publications which mentioned that almost every part of the tree is edible and very rich in its own nutritive and medicinal constituents. The tree is being used for many other purposes; even the bark of the tree represents an excellent anti fire fiber. In the present study the dormancy of seeds of *Adansonia digitata* L. was successfully broken up by pretreatment with full and half strength of hydrochloric, sulfuric and nitric acids for one and/or two hours. The obtained seedlings were used as a source of nodal segments explants for propagation in vitro. Regeneration of plantlets from the nodal segments of the obtained seedlings was carried out on MS culture medium using benzyl adenine as a cytokinin (from 2-10 mg/l) with or without the addition of naphthelene acetic acid (0.5 -1.0) as an auxin. After the obtained regenerants had reached suitable height and developed roots, they were acclimatized and transferred to soil successfully. The results obtained in this study may play a role in saving and spreading this tree in Yemen for local use and probably, improvement of country national income.

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كسر كمون بذور نبات أدانسونيا ديجيتاتا واستخلاف النبيتات من القطع الساقية العقديّة في الأنابيب**عزيزة مصلح تاج الدين****قسم البيولوجي – كلية العلوم – جامعة صنعاء - اليمن**

إن نبات أدانسونيا ديجيتاتا نبات معرض للخطر والإنقراض ليس فقط في البلدان التي أدخل إليها ولكن أيضا في موطنه الطبيعي نتيجة الحصاد الجائر وكمون البذور وضعف الاستخلاف الطبيعي وبطئ النمو. في اليمن توجد فقط ثلاث أشجار وفي الهند يوجد من 30-40 شجرة فقط. أدانسونيا ديجيتاتا (الذي يطلق عليها العامة في اليمن " شجرة الغريب") تعتبر واحدة من أطول الأشجار في العالم وتشير بعض الأبحاث إلى أنها من الممكن أن تعيش حتى 4000 سنة ويوجد العديد من الأبحاث التي تذكر أن كل جزء من الشجرة يمكن أن يؤكل وكل جزء غني بمكوناته الغذائية والدوائية. تستخدم الشجرة في العديد من الأغراض الأخرى وعلى سبيل المثال فإن قلف الشجرة يعتبر من الألياف المقاومة للنار. لقد تم في الدراسة الحالية كسر كمون بذور النبات بنجاح بالمعاملة بأحماض الهيدروكلوريك والكبريتيك والنيتريك لمدة ساعة أو ساعتين. لقد تم استخدام البادرات الناتجة من إنبات البذور في الأنابيب كمصدر للمنفضلات النباتية في صورة قطعاً ساقية عقديّة. لقد أمكن استيلاء خلفات نباتية من تلك القطع الساقية العقديّة في الأنابيب على وسط موراشسج وسكوج المزود بالبنزول أدينين منفردا كسيتوكينين (من 2 إلى 10 مج للتر) مع إضافة أو عدم إضافة نفتالين حامض الخليك كأوكسين (من نصف إلى واحد مج للتر). بعد وصول الخلفات في الأنابيب إلى الطول المناسب وتكوين الجذور كان من الممكن أقلمتها ونقلها إلى الأرض المستديمة. إن النتائج المتحصل عليها من هذه الدراسة ربما تلعب دورا في إنقاذ النبات واستزراع تلك الشجرة باليمن للإستخدام المحلي وربما لتحسين الدخل القومي للبلاد.